

Sequencing from single cells!

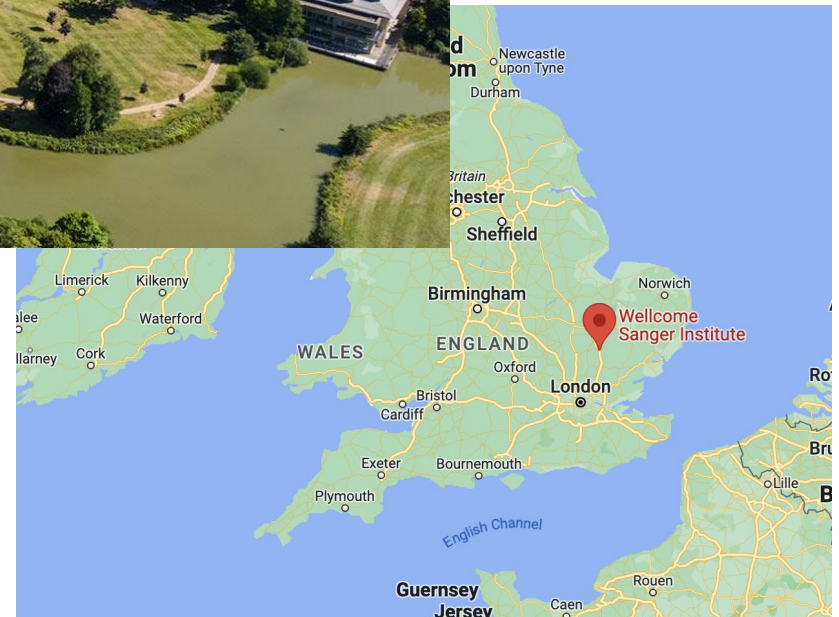
Lia Chappell

Wellcome Sanger Institute, Cambridge, UK

LC5@sanger.ac.uk

Twitter: @LiaVLChappell

I'm based the Sanger Institute



I grew up in Oxfordshire...

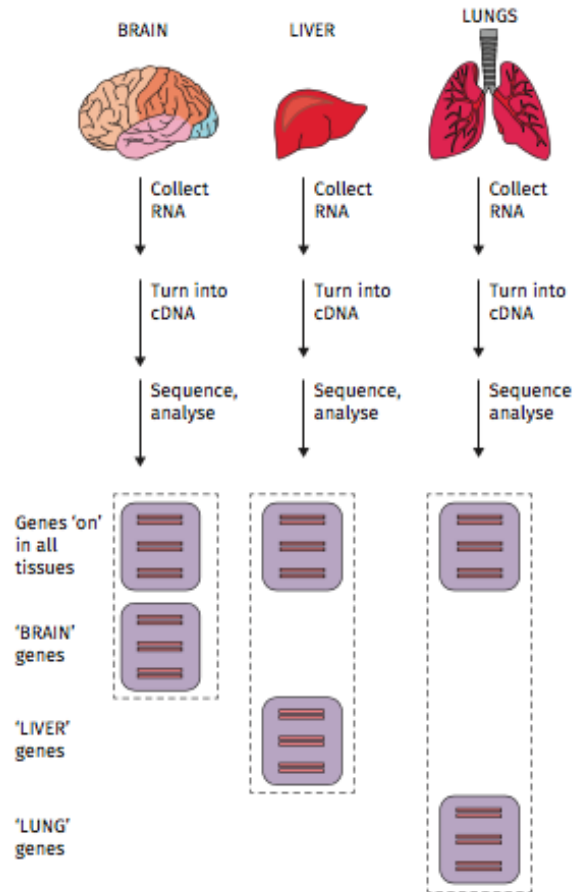


I'm from Oxfordshire

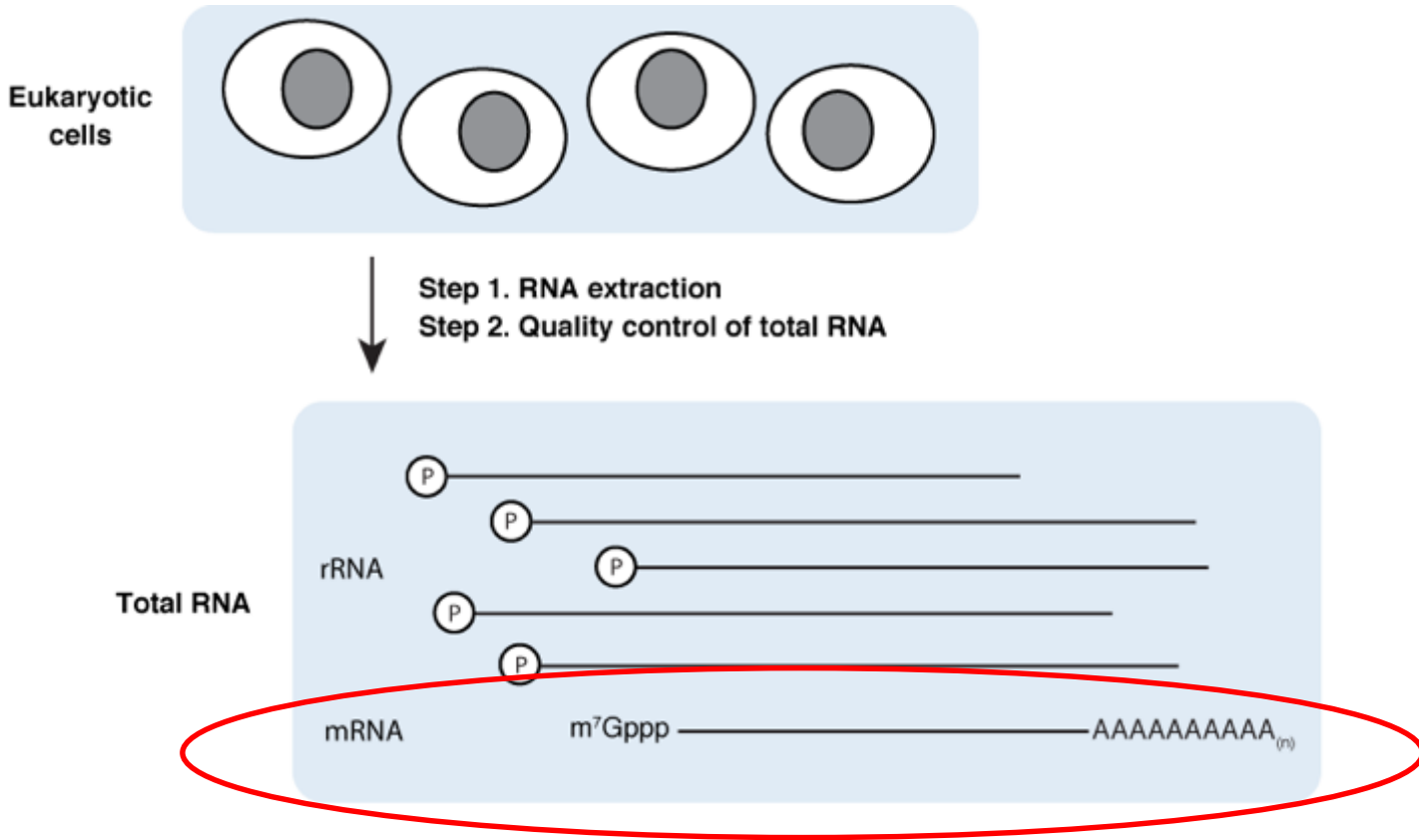


“Bulk” RNA-seq

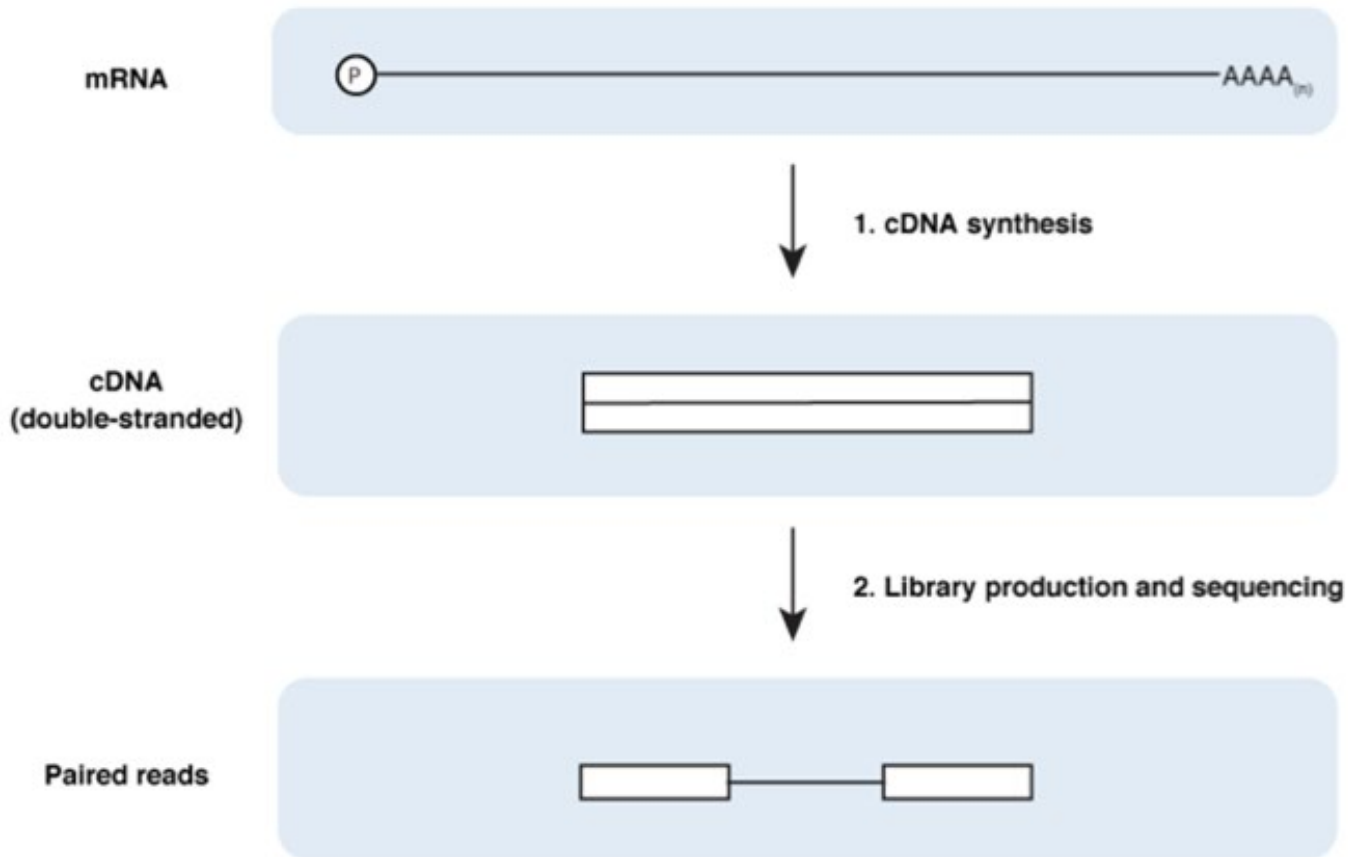
RNA-seq measures “active” genes with sequencing



RNA



From RNA to RNA-seq



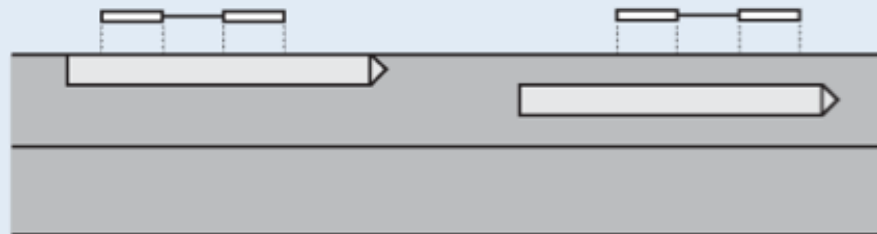
From RNA to RNA-seq

Paired reads



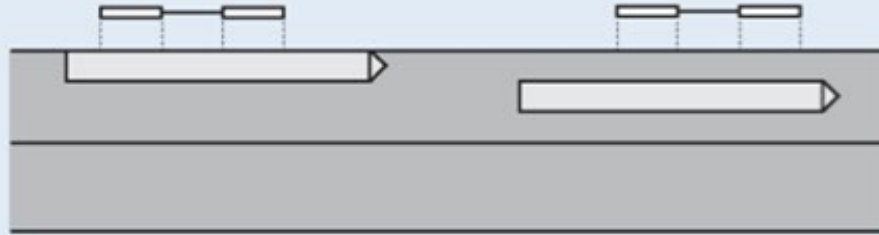
3. Read mapping

Reads mapped to the genome



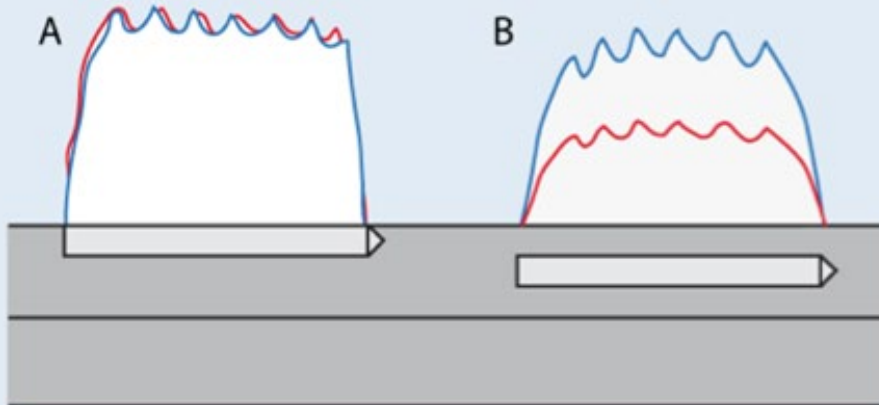
From RNA to RNA-seq

Reads mapped to the genome



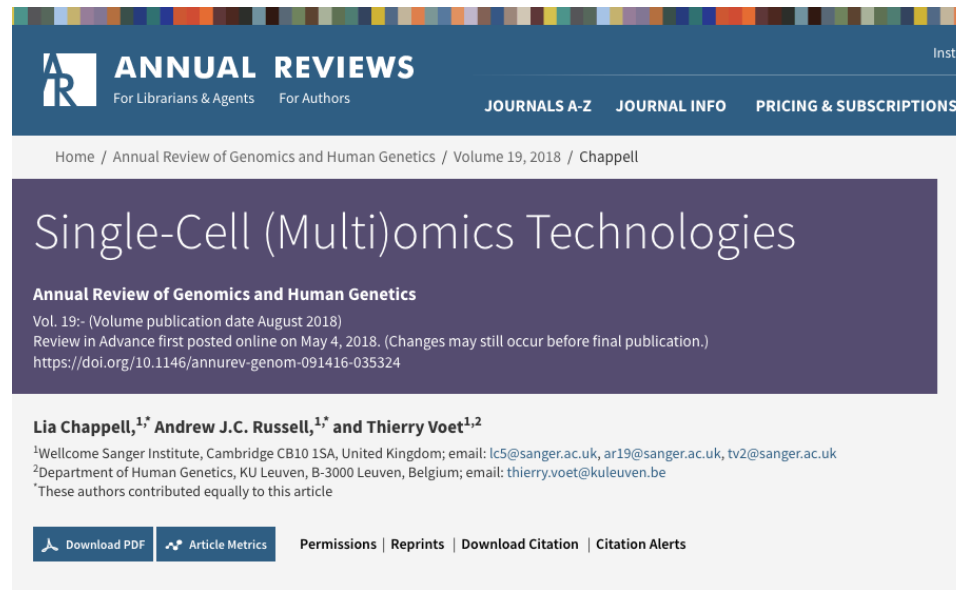
4. Generate coverage plots

Coverage plots visualise differential gene expression between biological conditions



Sequencing from single cells: why and how?

Too long and full of too many technical details.....



The screenshot shows the top portion of a web page for an Annual Review article. At the top is a dark blue header with the 'AR' logo and the text 'ANNUAL REVIEWS For Librarians & Agents For Authors'. Navigation links include 'JOURNALS A-Z', 'JOURNAL INFO', and 'PRICING & SUBSCRIPTIONS'. Below the header is a breadcrumb trail: 'Home / Annual Review of Genomics and Human Genetics / Volume 19, 2018 / Chappell'. The main title 'Single-Cell (Multi)omics Technologies' is displayed in a large, light-colored font against a dark purple background. Below the title, it identifies the journal as 'Annual Review of Genomics and Human Genetics', Volume 19, published in August 2018, with a review in advance posted online on May 4, 2018. The authors are listed as Lia Chappell, Andrew J.C. Russell, and Thierry Voet. A note states that these authors contributed equally to the article. At the bottom of this section are buttons for 'Download PDF', 'Article Metrics', 'Permissions', 'Reprints', 'Download Citation', and 'Citation Alerts'.



Andy Russell



Thierry Voet

Abstract

Single-cell multiomics technologies typically measure multiple types of molecule from the same individual cell, enabling more profound biological insight than can be inferred by analyzing each molecular layer from separate cells. These single-cell multiomics technologies can reveal cellular heterogeneity at multiple molecular layers within a population of cells and reveal how this variation is coupled or uncoupled between the captured omic layers. The data sets generated by these techniques have the potential to enable a deeper understanding of the key biological processes and mechanisms driving cellular heterogeneity and how they are linked with normal development and aging as well as disease etiology. This review details both established and novel single-cell mono- and multiomics technologies and considers their limitations, applications, and likely future development



Review in Advance first posted on
 May 4, 2018. (Changes may still
 occur before final publication.)

Table 1 Single-cell multiomics technologies

Technology	Genomic layer	Chromatin accessibility epigenetic layer	CpG methylation epigenetic layer	Transcriptomic layer	Protein layer	Strategy	Cell isolation ^a	Cell throughput	Automation	Constituent methods
DR-seq (31)	Genome	—	—	Transcriptome	—	Preamplification and tagging of DNA and RNA followed by splitting	Mouth pipette	Low	No	Modified CEL-seq (59) and modified MALBAC (161)
G&T-seq (92, 94)	Genome	—	—	Transcriptome	—	Separation (DNA and polyadenylated mRNA)	FACS	Medium	Yes	Modified Smart-seq2 (112, 113) and PicoPLEX WGA or MDA WGA (92, 94)
scTrio-seq (62)	CNVs (from scRRBS data)	—	Reduced-representation DNA CpG methylation	Transcriptome	—	Separation (nucleus and cytoplasm) followed by bisulfite conversion	Mouth pipette	Low	No	scRNA-seq method of Tang et al. (143) and scRRBS (54)
scMT-seq (64)	SNPs (from scRRBS and scRNA-seq data)	—	Reduced-representation DNA CpG methylation	Transcriptome	—	Separation (nucleus and cytoplasm)	Microcapillary pipette	Low	Partial	Modified Smart-seq2 protocol (112, 113) and modified scRRBS (54)
scGEM (20)	Targeted genotyping (Sanger and next-generation sequencing)	—	Targeted DNA CpG methylation (qPCR)	Targeted (RT-qPCR)	—	Restriction digestion, preamplification, and splitting	Fluidigm C1	Medium	Yes	Modified SCRAM (21)
scM&T-seq (5)	—	—	DNA CpG methylation	Transcriptome	—	Separation (DNA and polyadenylated mRNA) followed by bisulfite conversion	FACS	Medium	Yes	Modified G&T-seq (92, 94) and modified scBS-seq (24, 131)

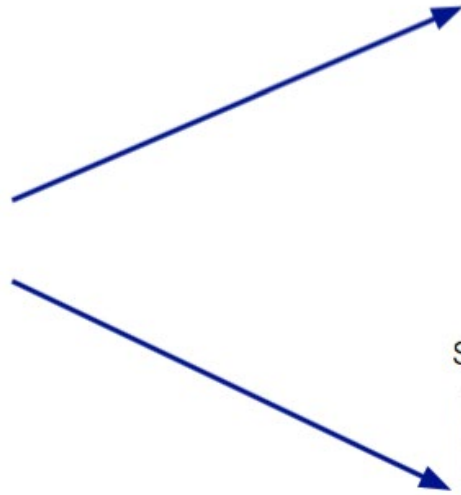
(Continued)

Why?



Bulk population based methods





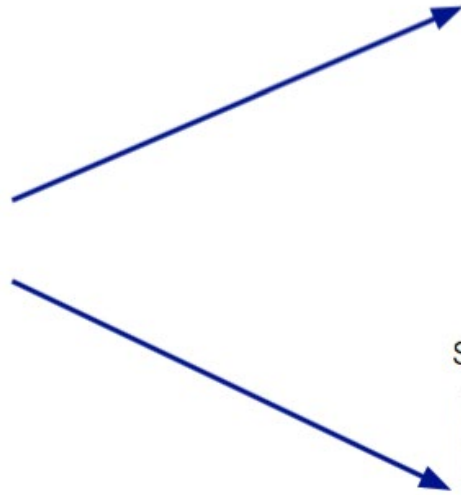
Bulk population based methods



Single Cell Methods



The Art of Clean Up, Ursus Wehrli, Kimberly Vardeman



Bulk population based methods



Single Cell Methods



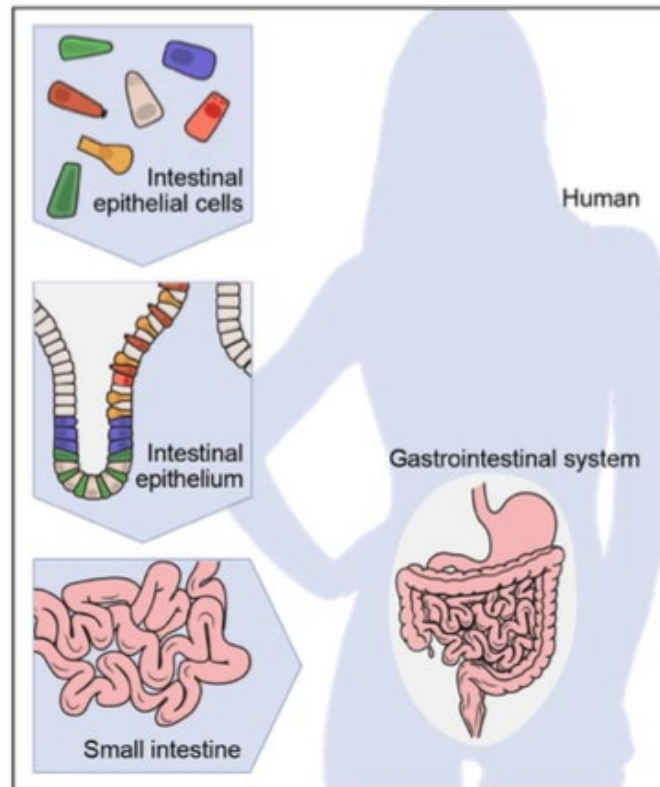
Spatial methods



The Art of Clean Up, Ursus Wehrli, Kimberly Vardeman




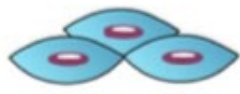
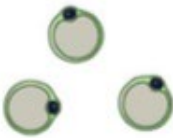




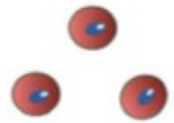
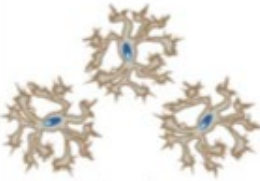
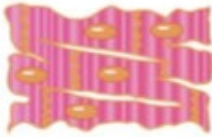
Human Cell Atlas

A “google maps” of human anatomy

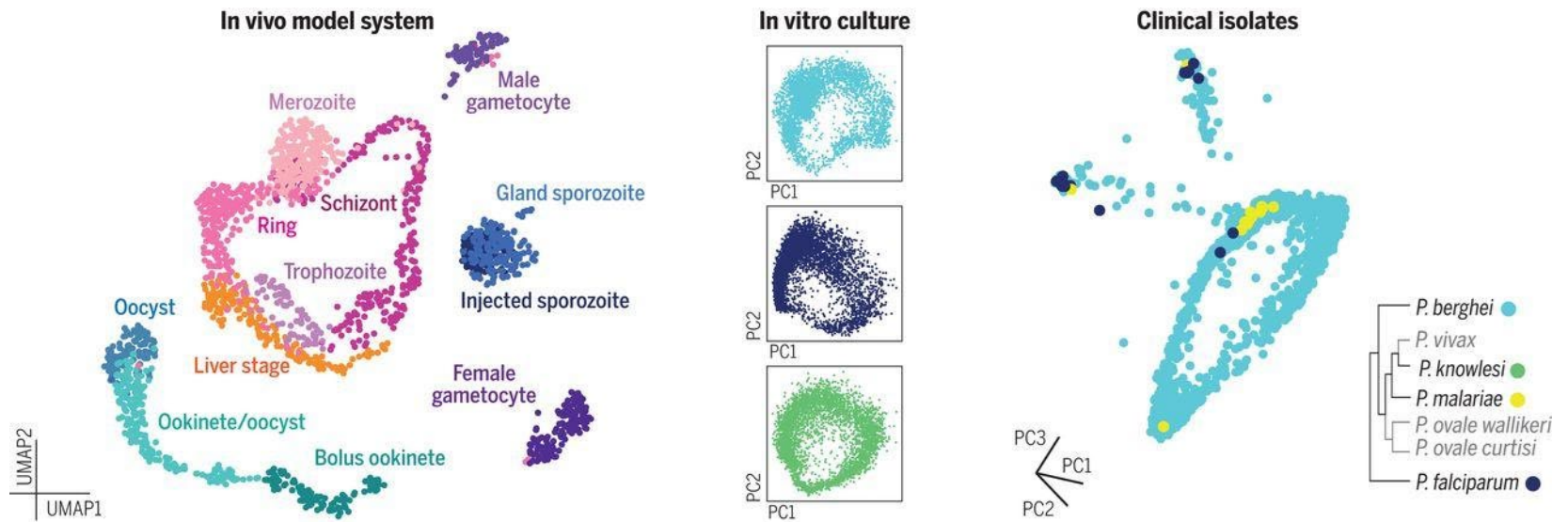


Human Cell Atlas

Goal : A periodic table of our cells

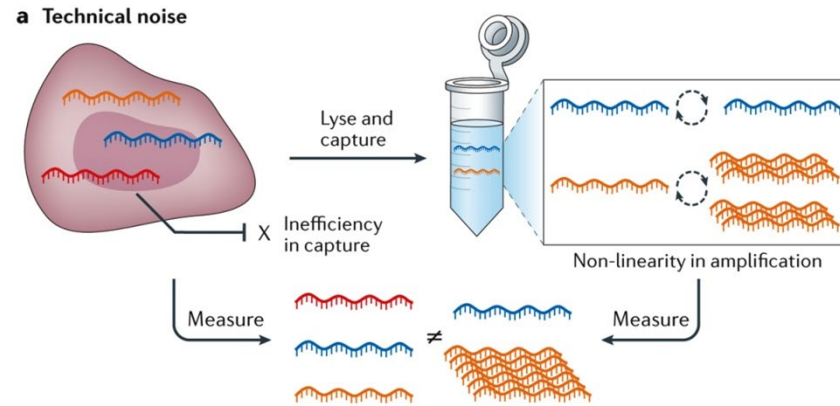
Skin Cells	Immune Cells	Brain Cells	Muscle Cells
 Fibroblasts	 Megakaryocytes	 Neurons	 Smooth Muscle
 Adipocytes	 Dendritic Cells	 Ependymal Cells	 Skeletal Muscle
 Epithelial Cells	 T Cells	 Astrocytes	 Cardiac Muscle

Malaria Cell Atlas



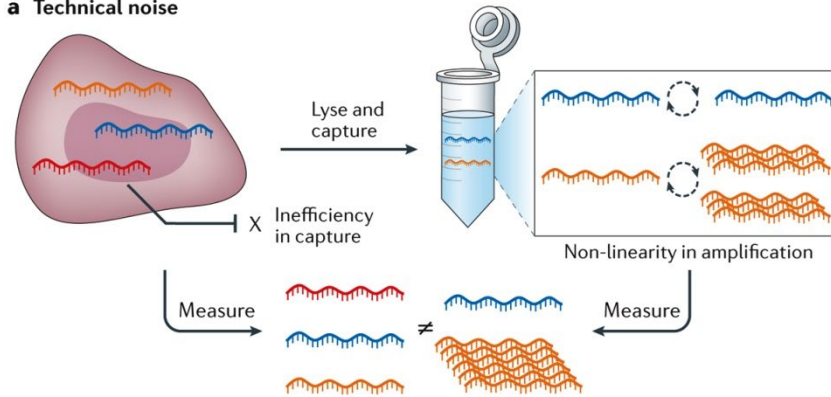
Virginia M. Howick et al. *Science* 2019;365:eaaw2619

Noise in scRNA-seq

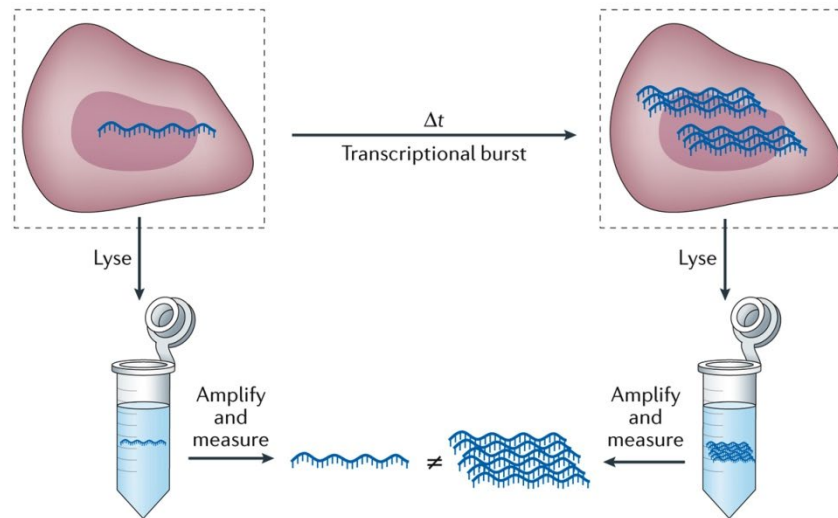


Noise in scRNA-seq

a Technical noise



b Intrinsic noise



How?

FACS machine



Usually in plates



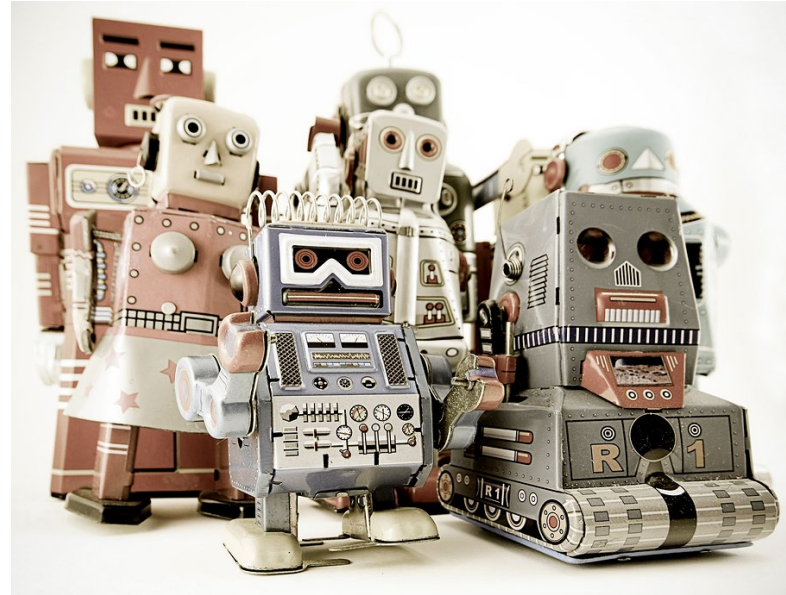
Single cell RNA-seq: plate format



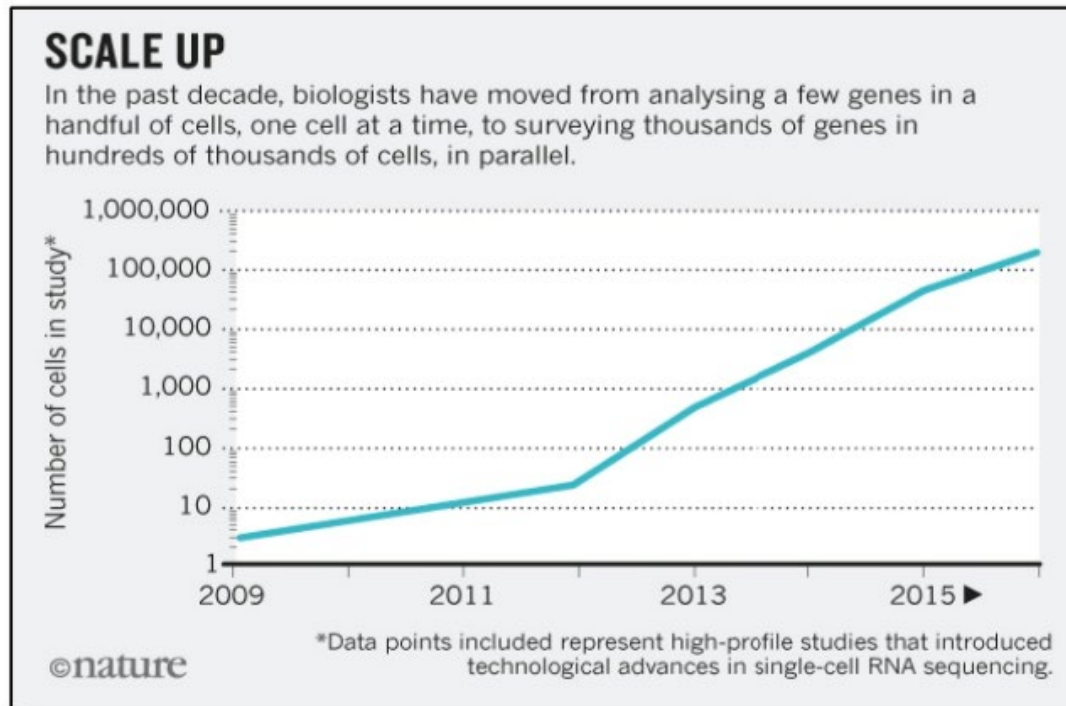
Single cell RNA-seq: plate format



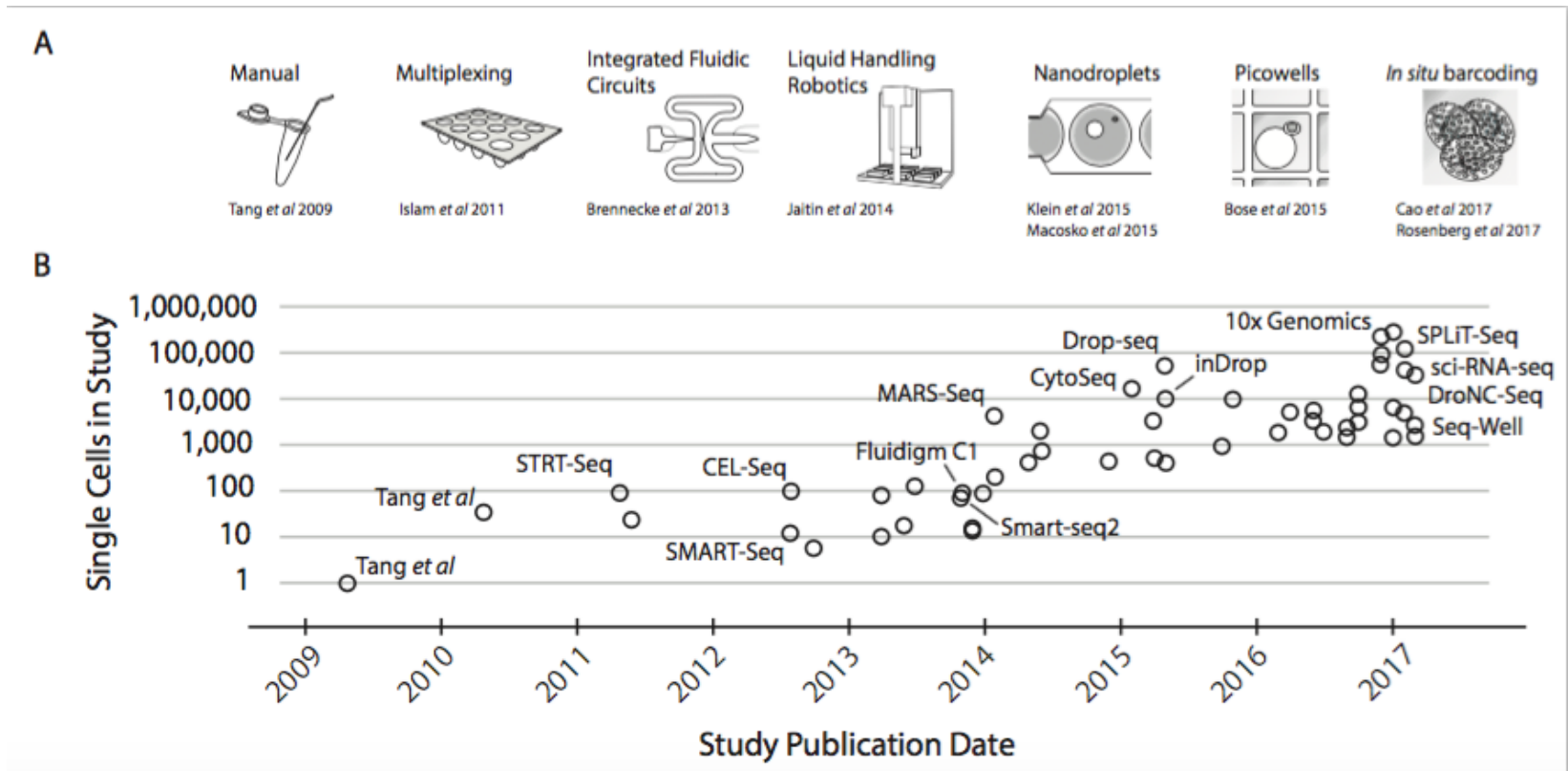
Single cell RNA-seq: plate format



A trend for increasing scale...

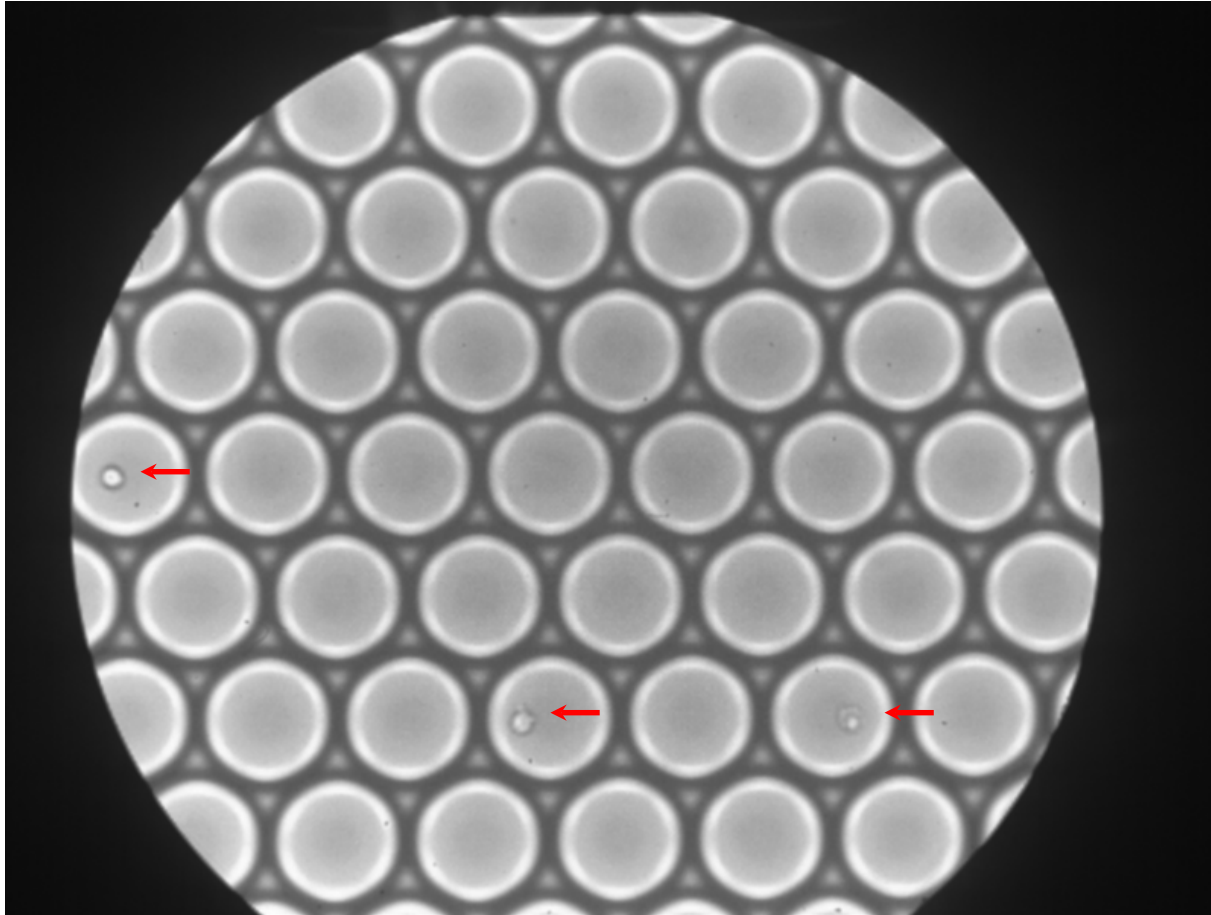


A trend for increasing scale...

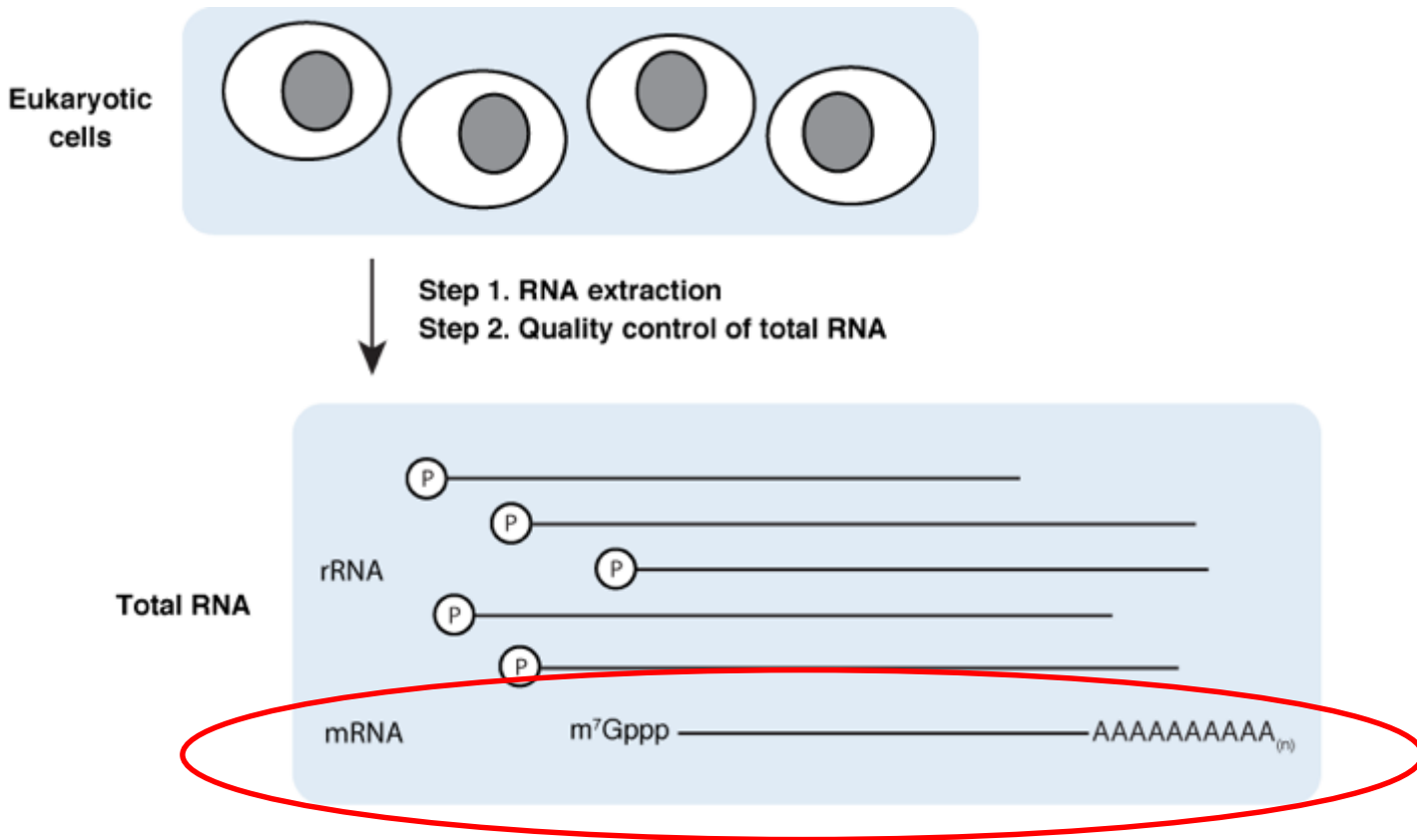


[Valentine Svensson](#), [Roser Vento-Tormo](#), [Sarah A Teichmann](#)

Another way: droplets

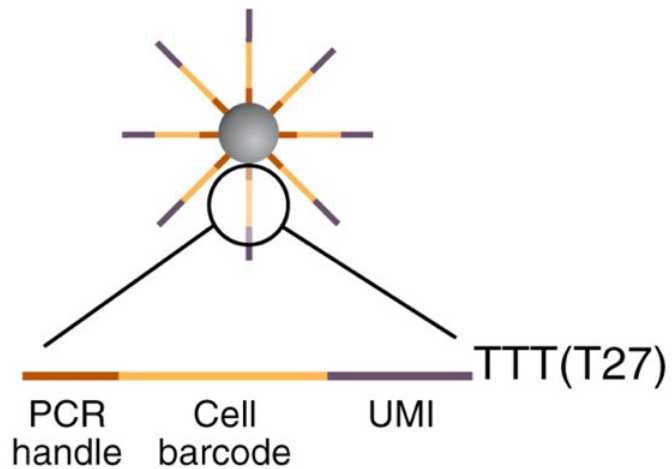


mRNA - a reminder



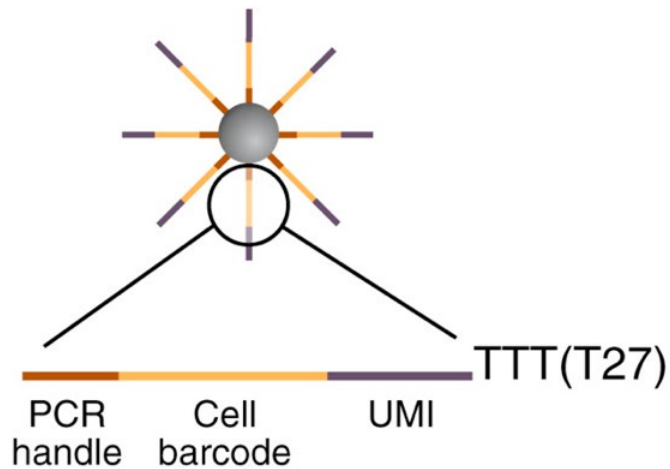
Barcoded beads...

Barcoded beads

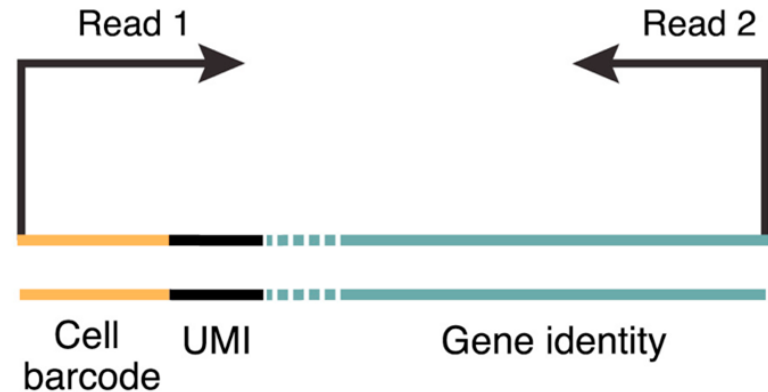


Barcoded beads...

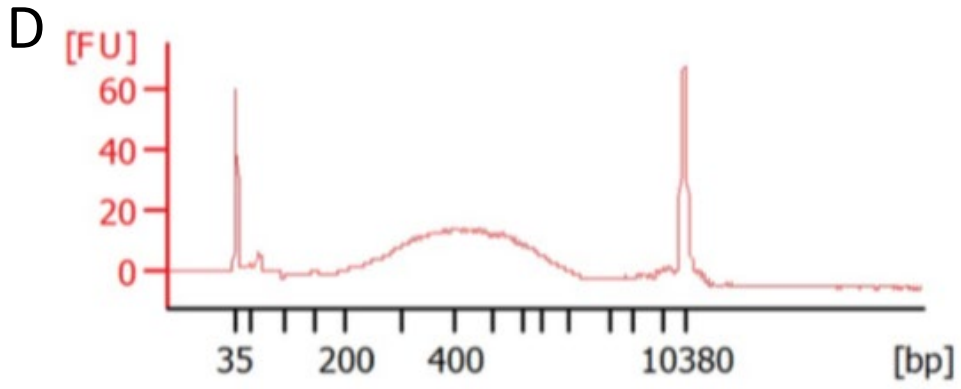
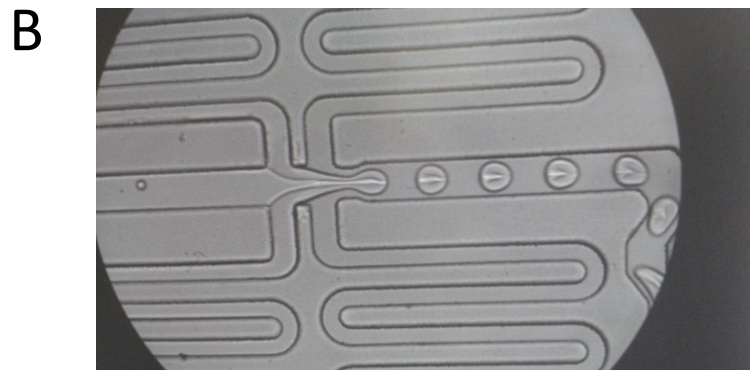
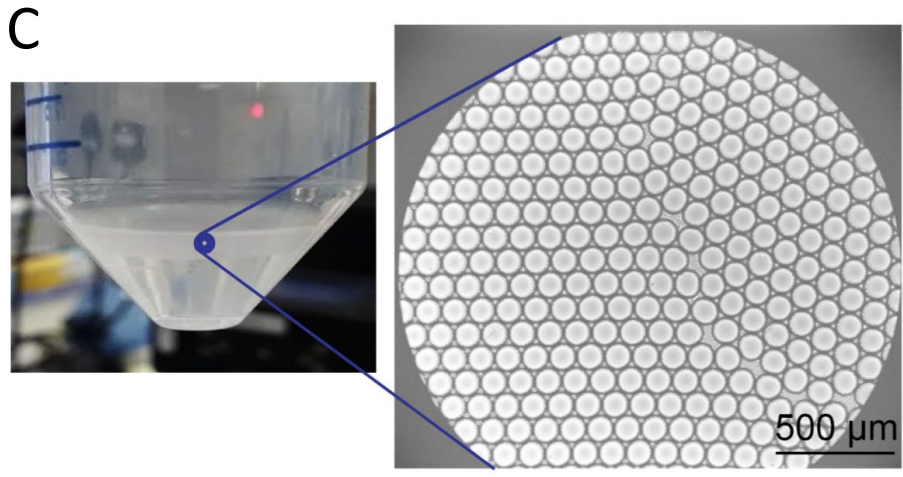
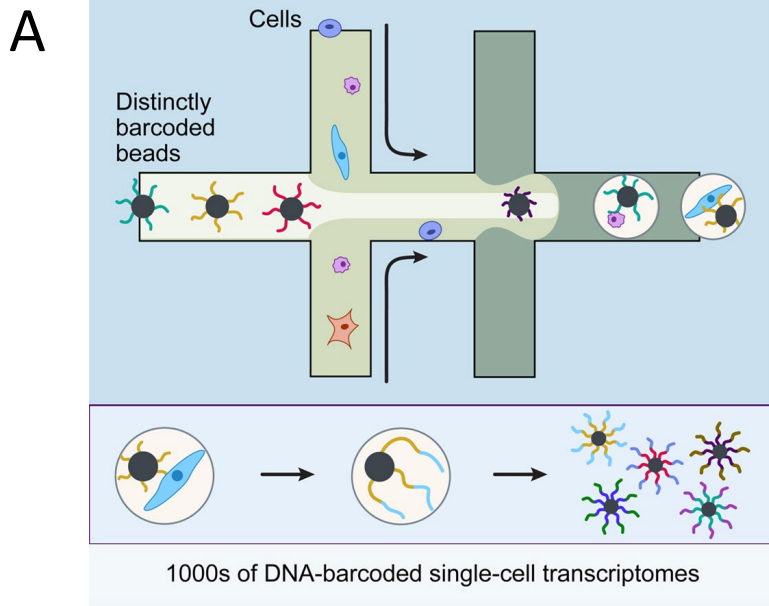
Barcoded beads



Sequencing reads



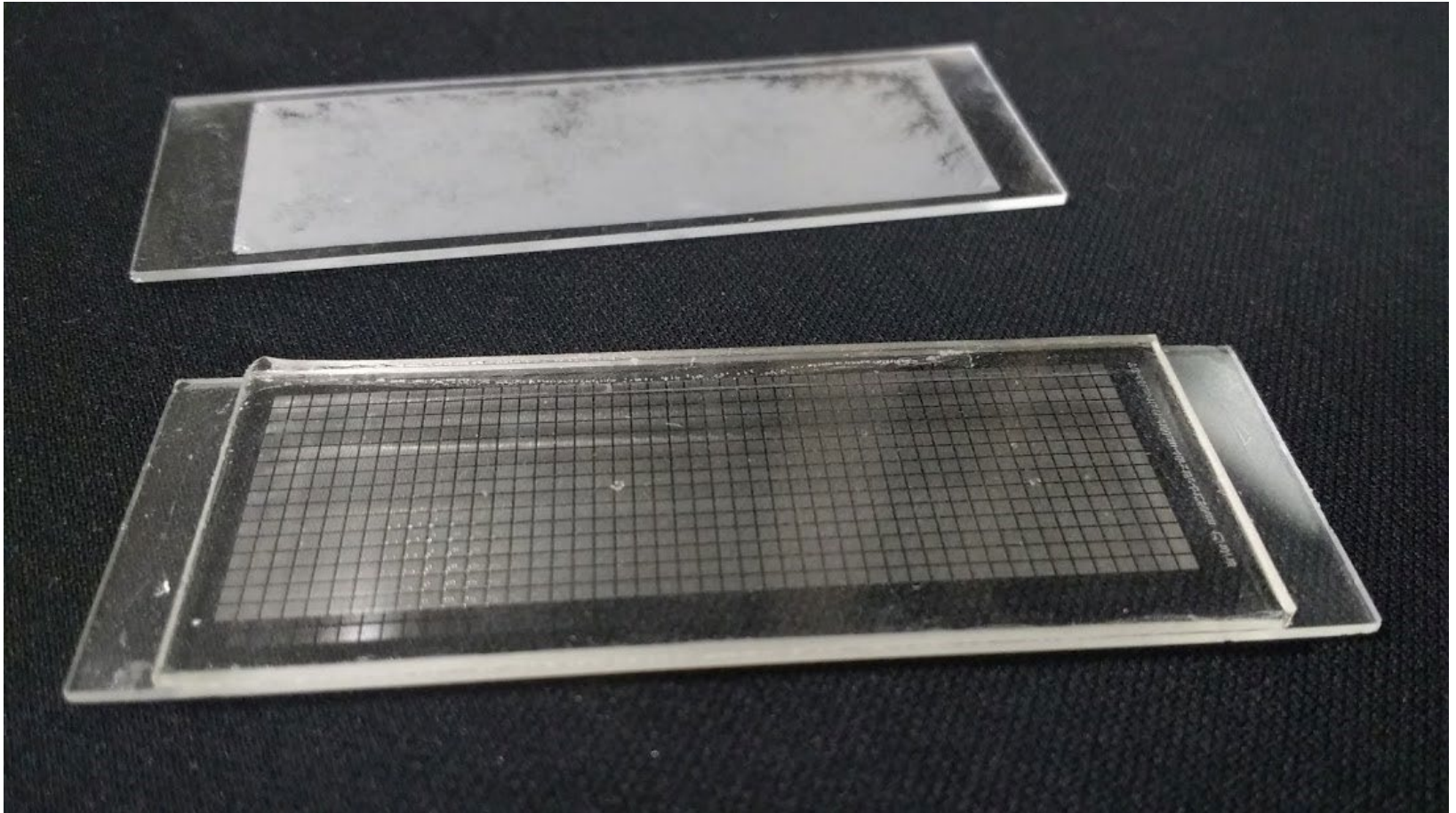
Drop-seq



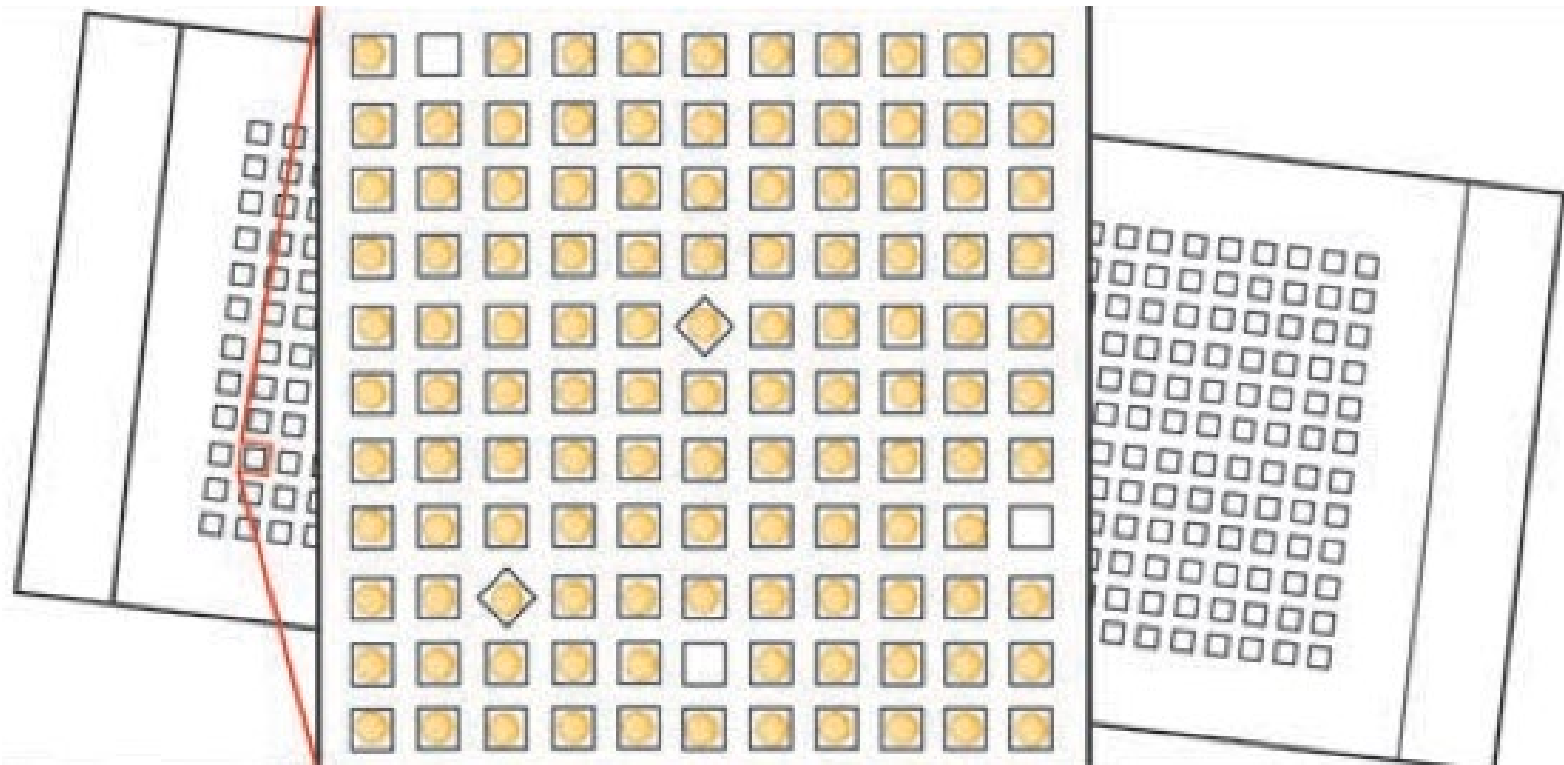
10X “black box”



Nanowells: e.g. Seq-Well



Beads fit into wells...

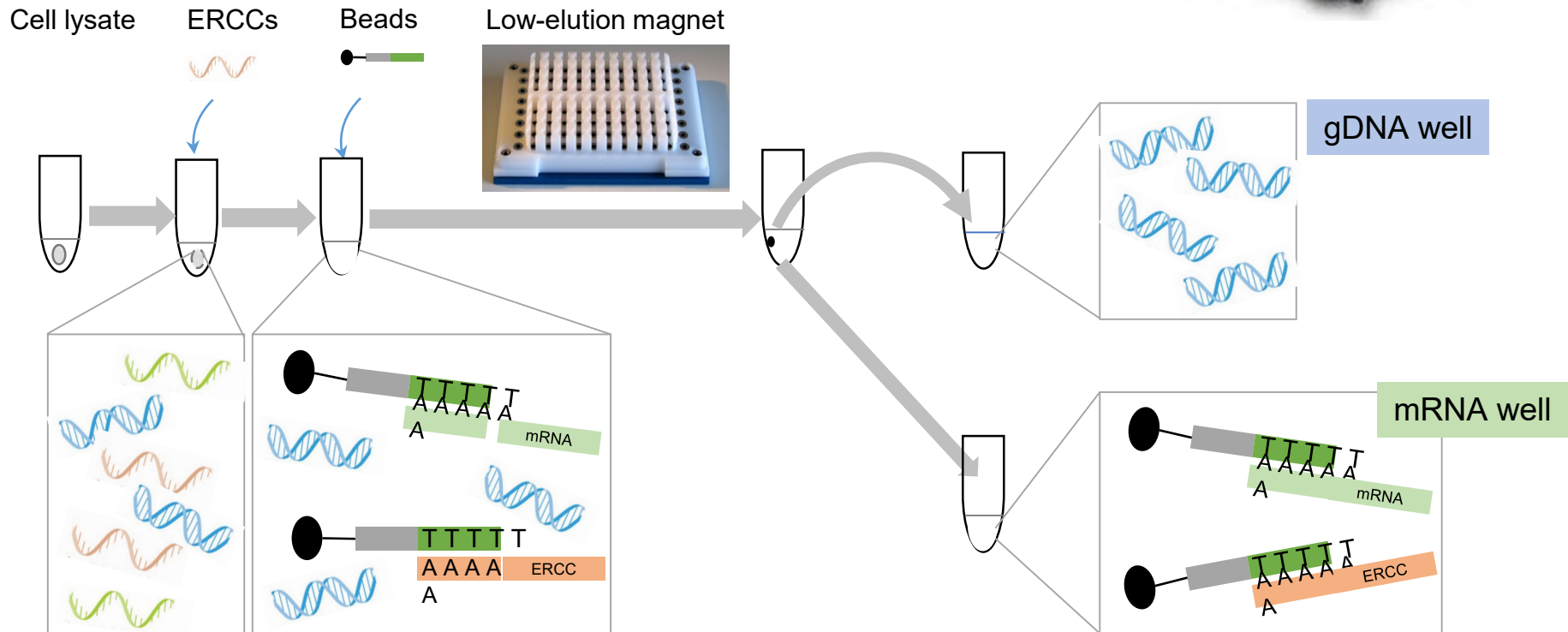


Multiple layers from
the same cell

Single-cell G&T-seq (Voet group)



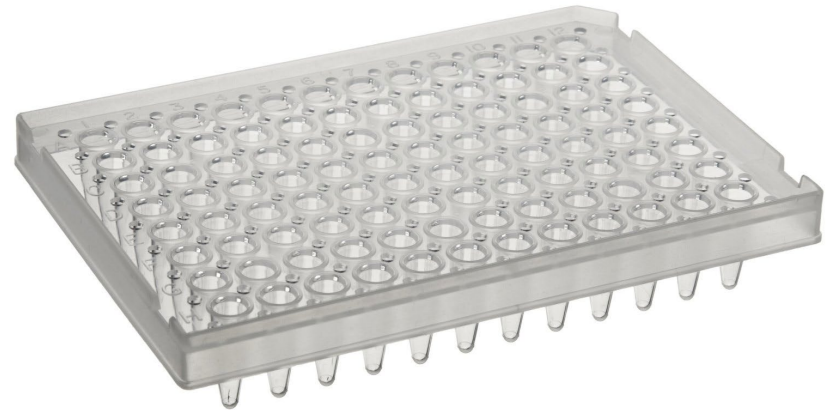
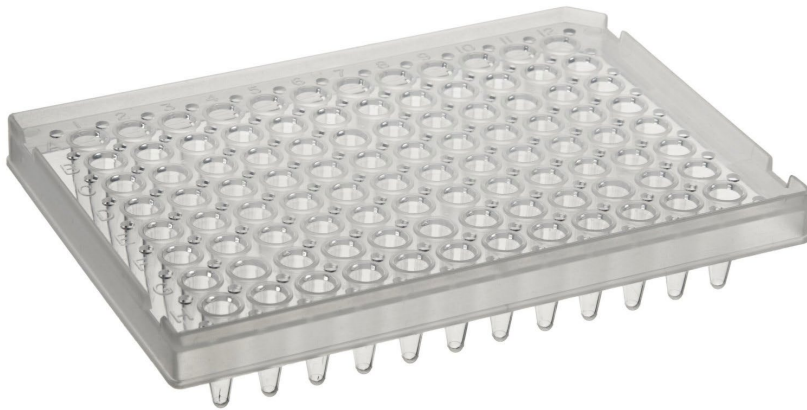
In plate format,
automated on
robotic platforms



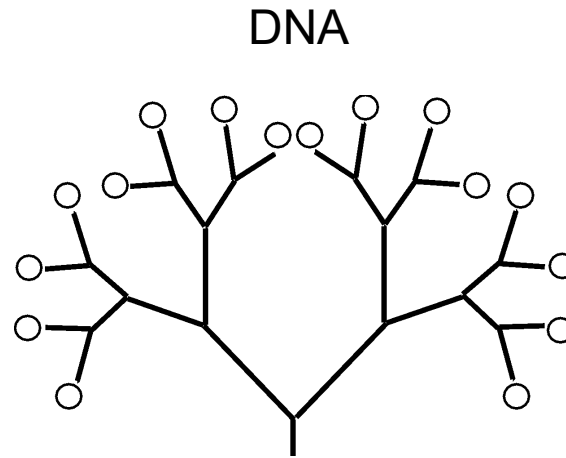
Relationship between RNA and DNA maintained by copying plate layout for both layers

RNA

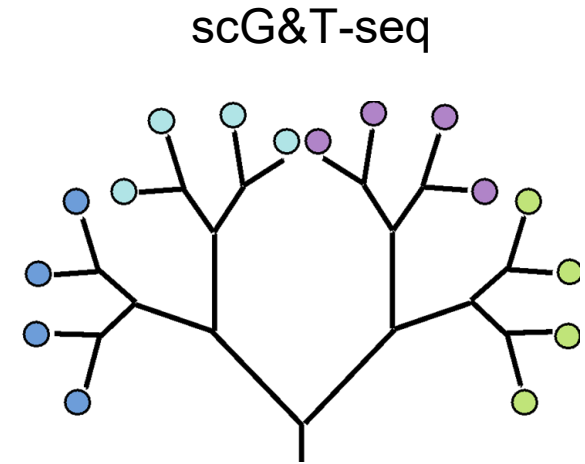
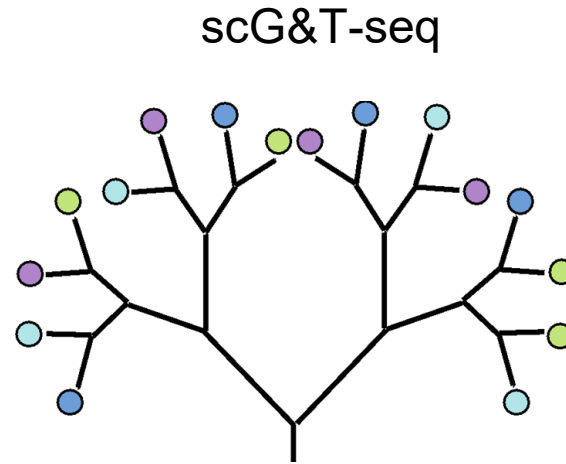
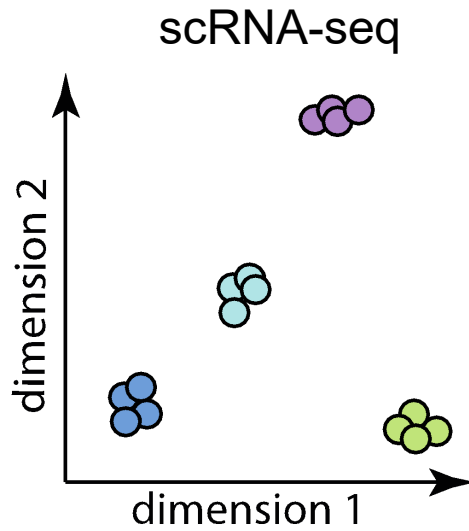
DNA



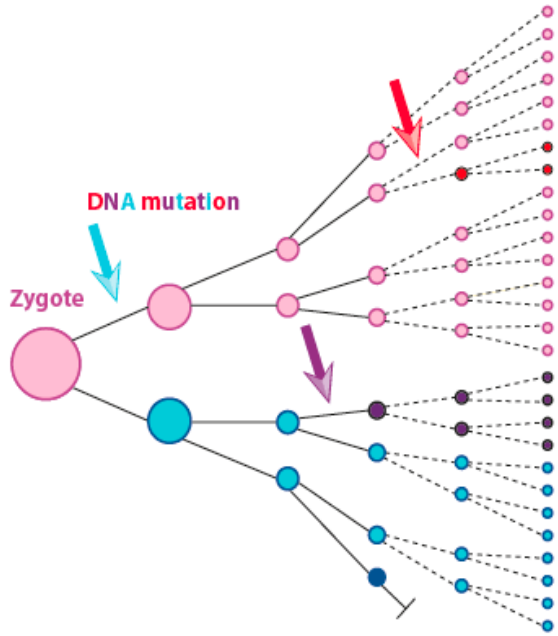
Genotype-phenotype correlation at single cell level



Genotype-phenotype correlation at single cell level

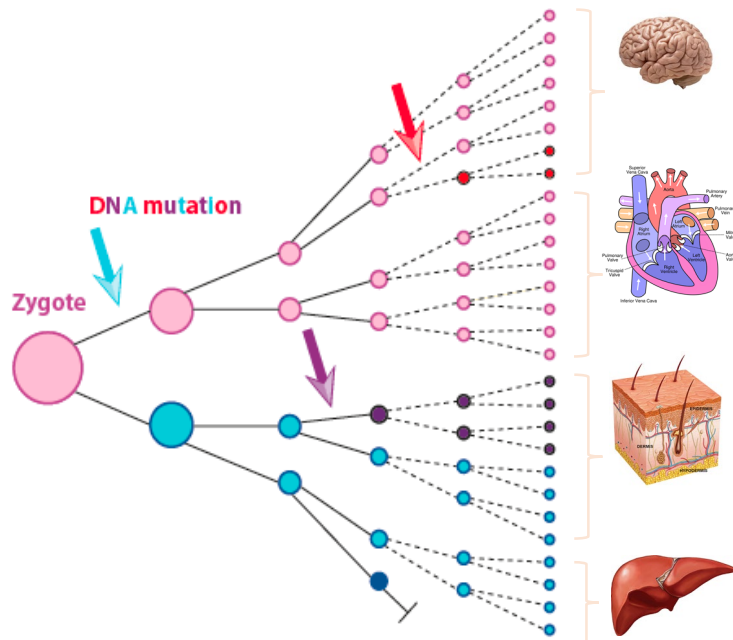


One human, multiple genomes



One human, multiple genomes

... as a means to study cellular architectures of human organs



- The relative contribution of embryonic cells to tissues/organs;
- The 'noise' in early embryonic development between individuals;
- Developmental and cellular architectures of organs:
 - clonal structures
 - amount of stem cells contributing to functional units
 - differentiation trajectories available to given adult stem cell populations;
- Cell lineage perturbed in diseased tissues/organs
- Nature and role of somatic mutation in phenotypic variation, aging and disease

Another dimension:
spatial

Modular valve positioner 1
+reagents

Modular valve positioner 2
+reagents

Scope

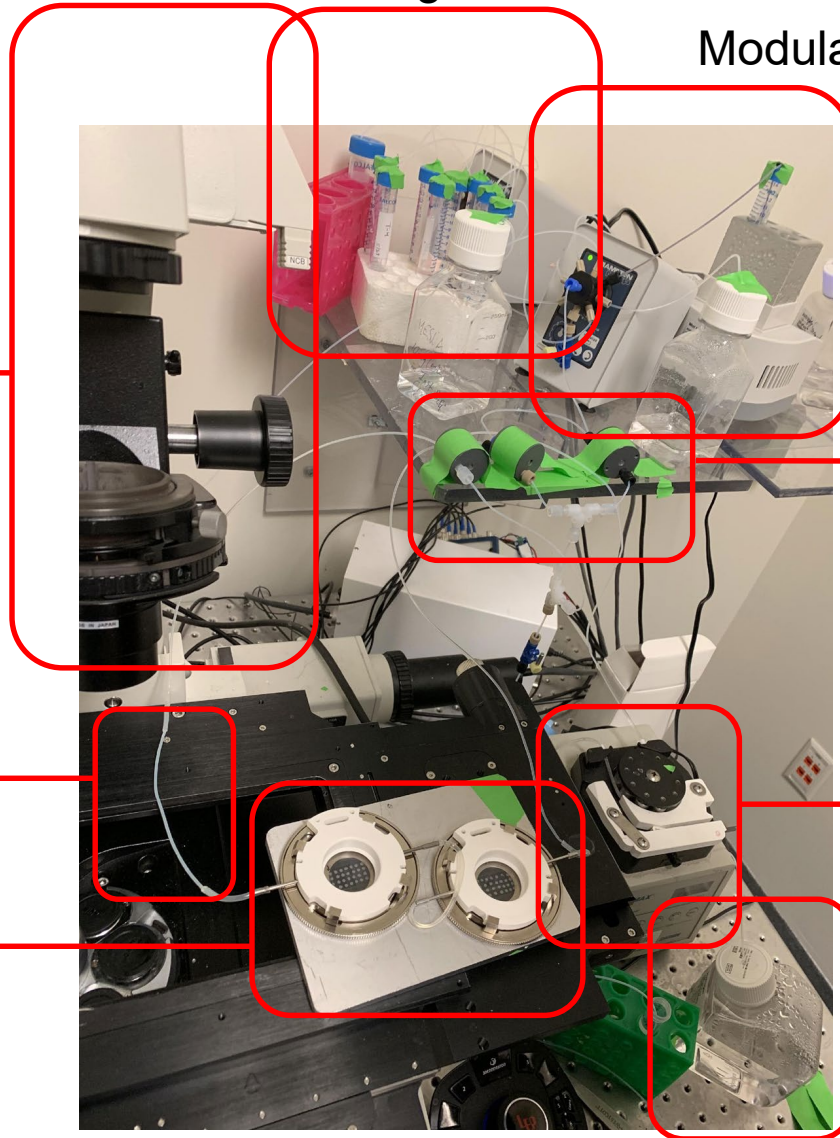
Bubble traps

Tubing

Peristaltic pump

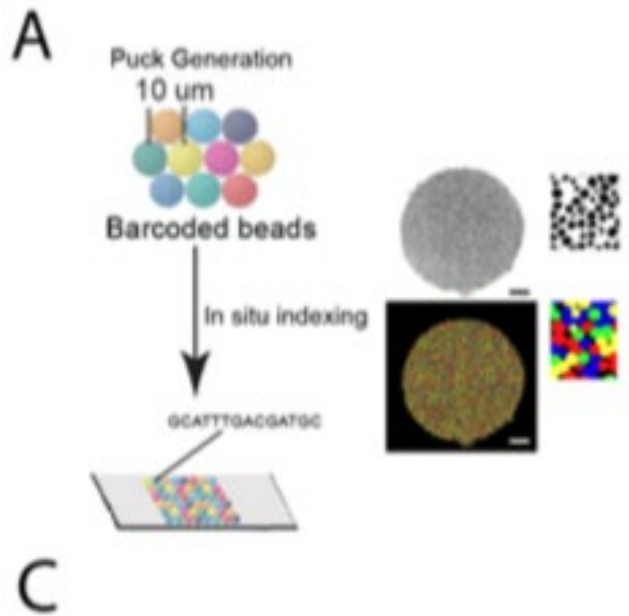
Flow cells

Waste



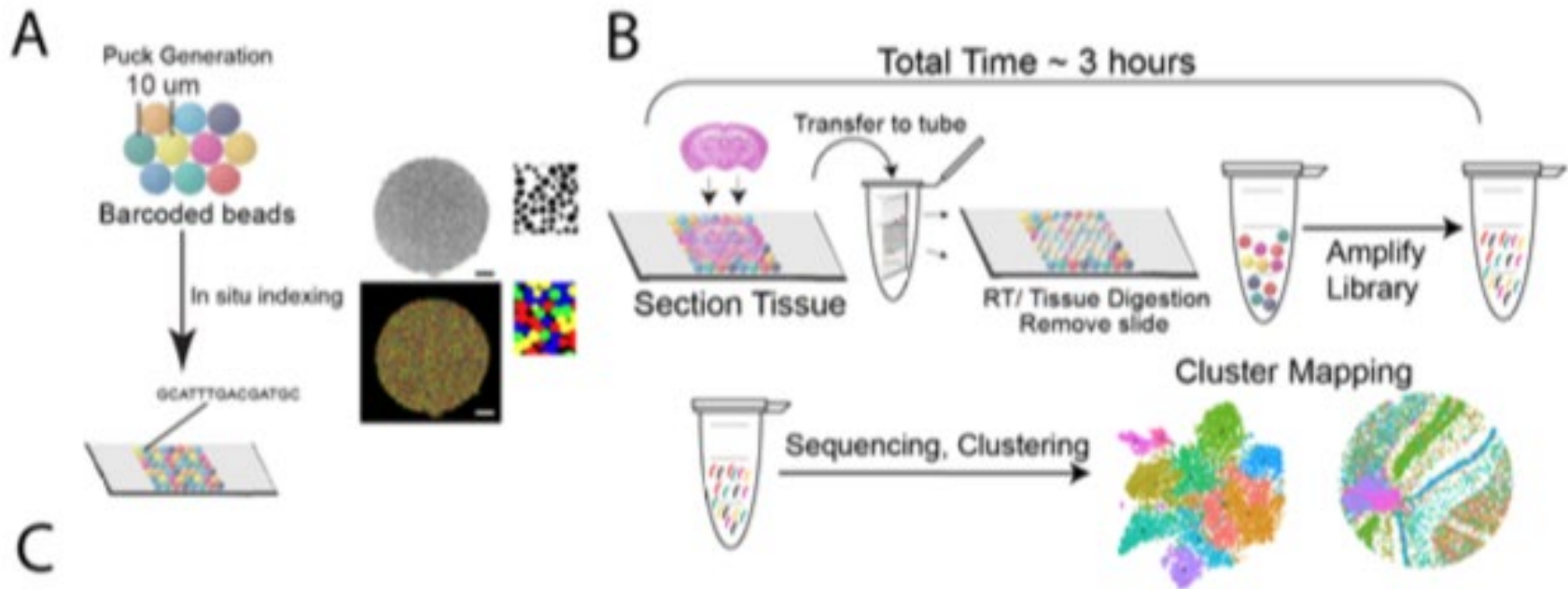
Slide-seq

- each bead barcode has known spatial location



Slide-seq

- each bead barcode has known spatial location



Slide-seq

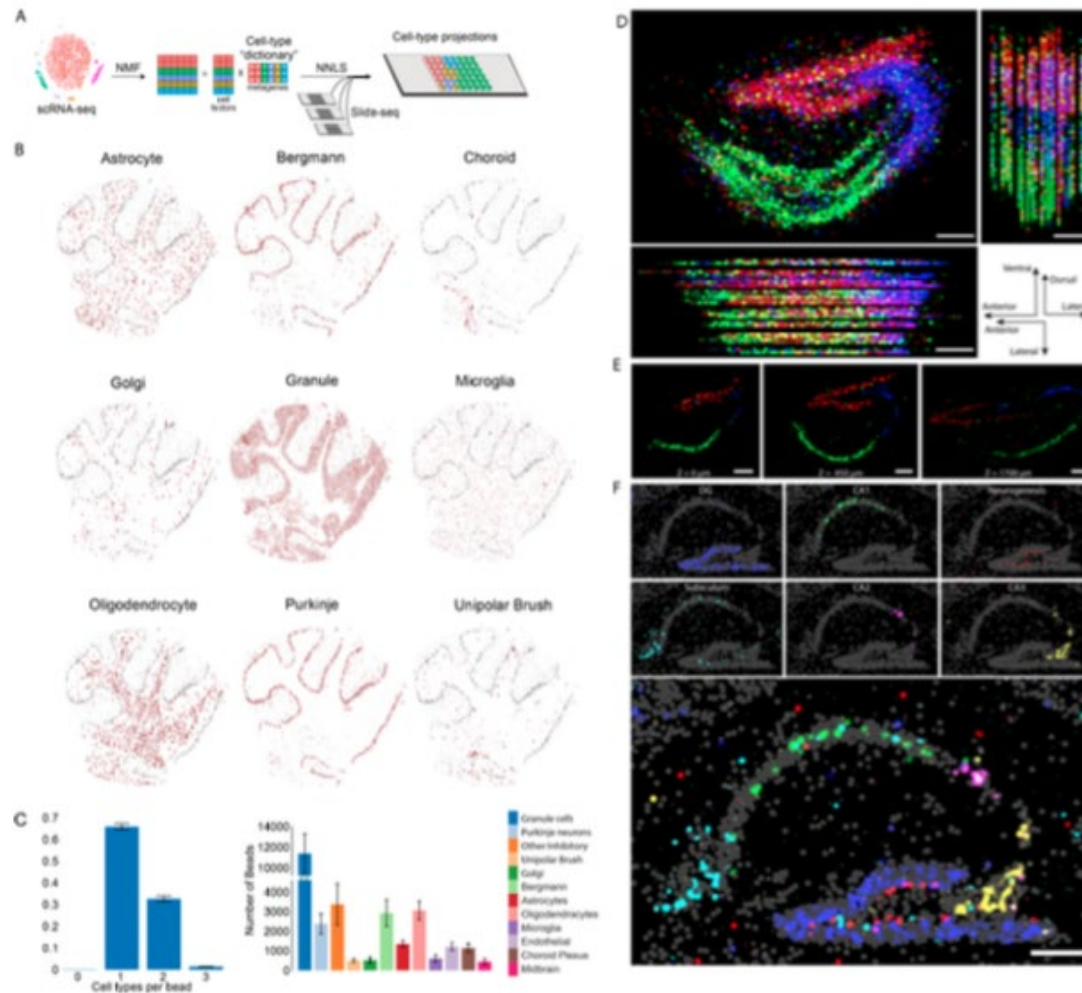
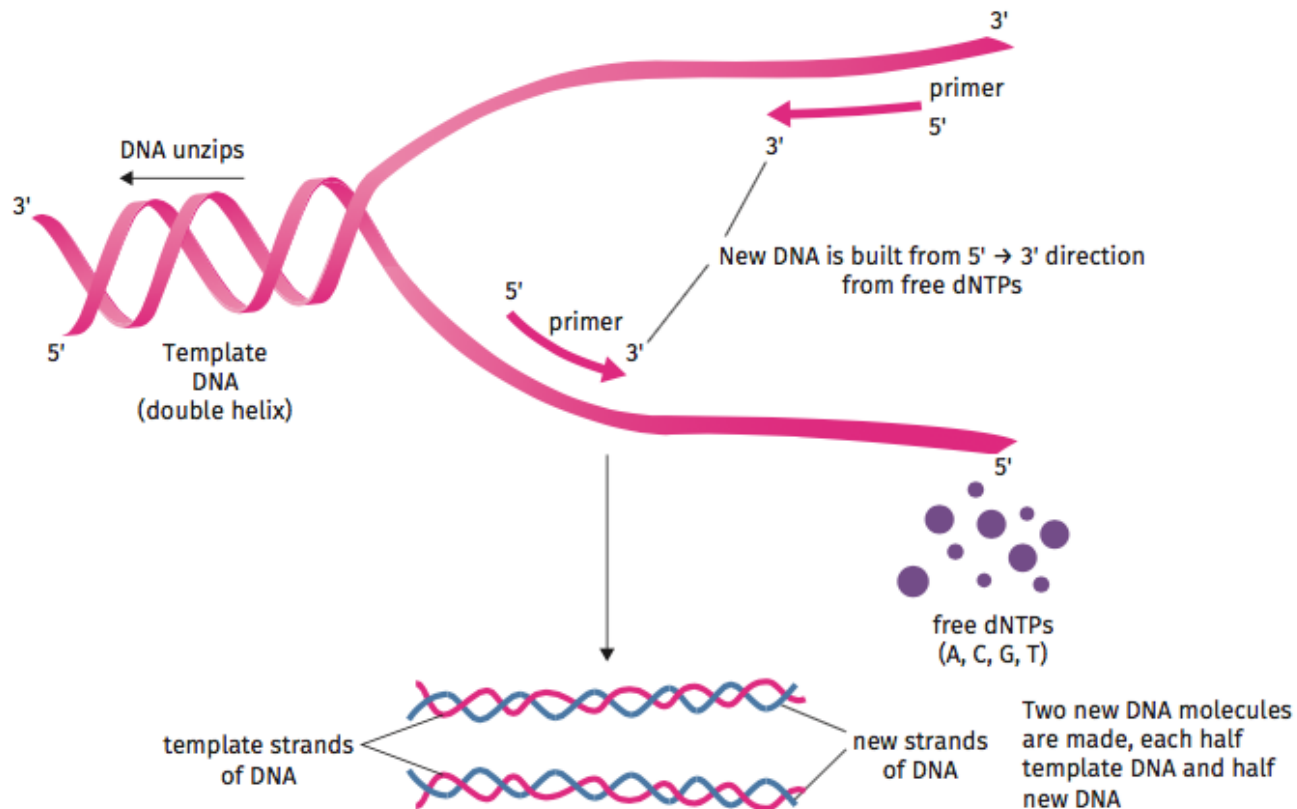


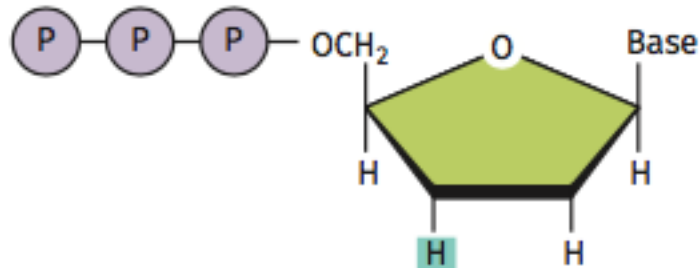
Figure 2: Localization of cell types in cerebellum and hippocampus using Slide-seq. (A)

Thank you for listening!
:)

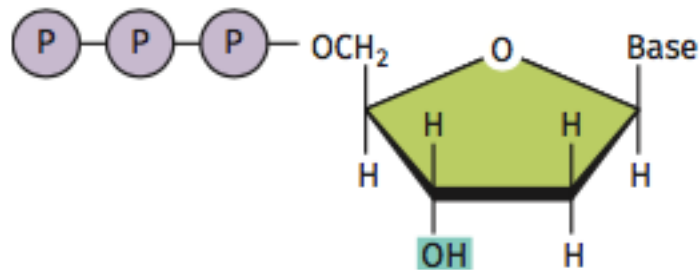
DNA replication: the key principles that underlie the tech



Nucleotide building blocks

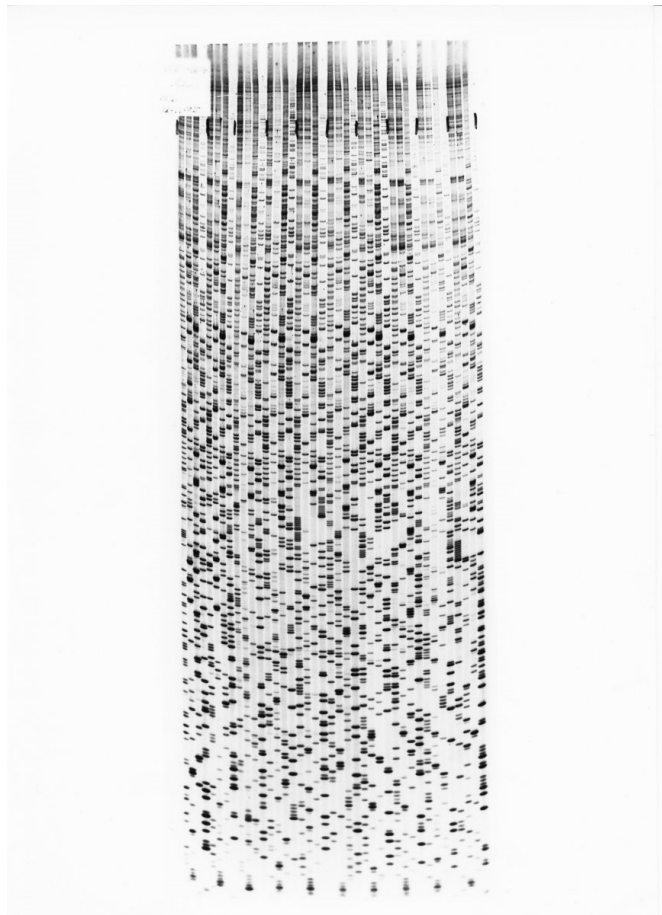


Dideoxynucleotide (ddNTP)

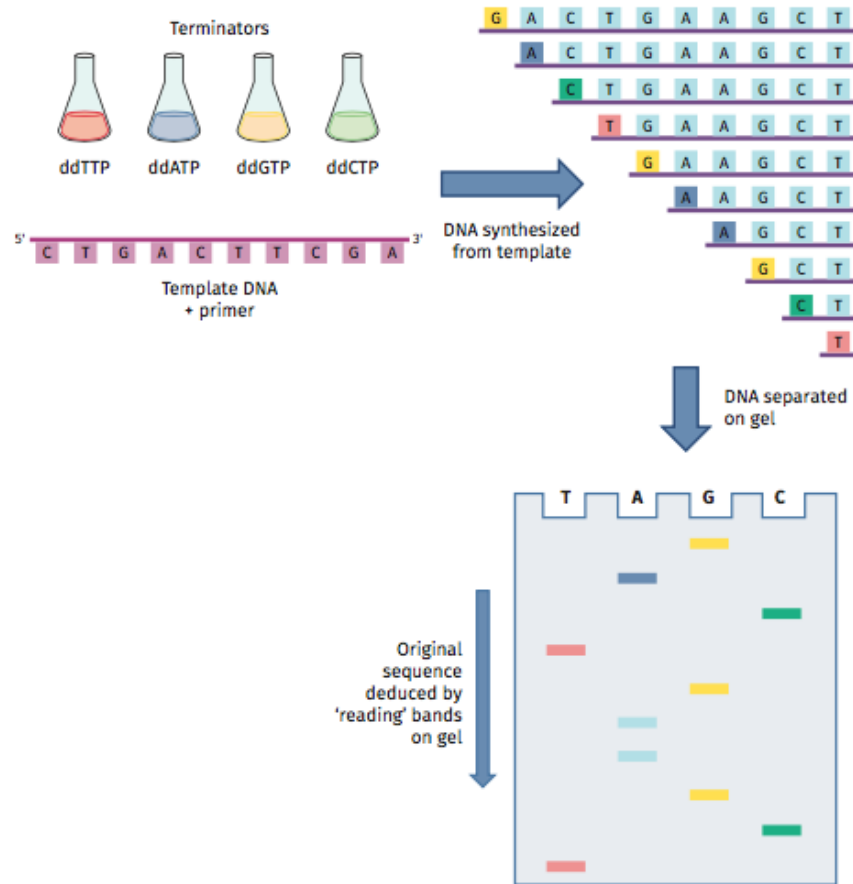


Deoxynucleotide (dNTP)

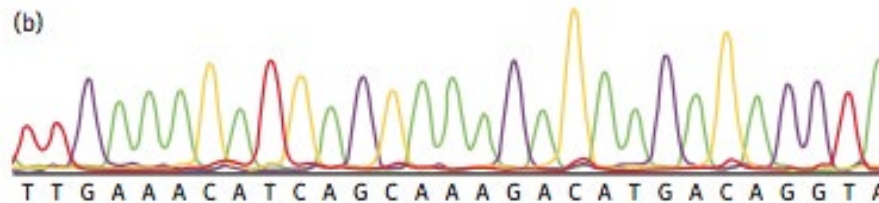
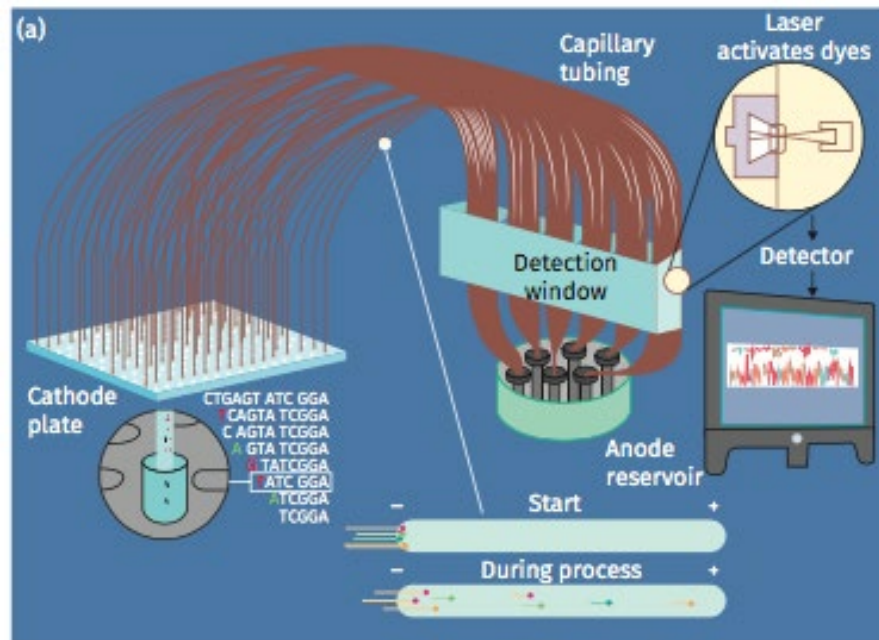
Sanger sequencing



Sanger sequencing



Capillary sequencing



“Next gen” sequencing: Illumina

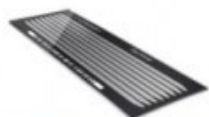


Illumina “flow cells”

HiSeq 2000
New flow cell design

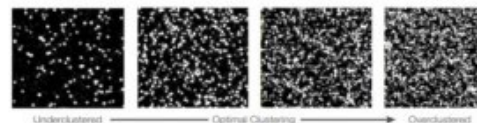
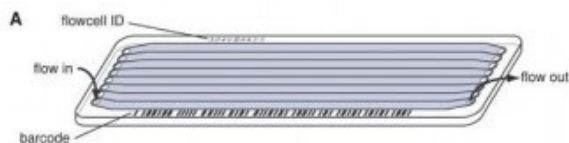
LARGER, DUAL-SURFACE ENABLED
->5x increase in imaging area
Retains 8 lane format

Compatible with cBot



Cluster density
750-850/mm²

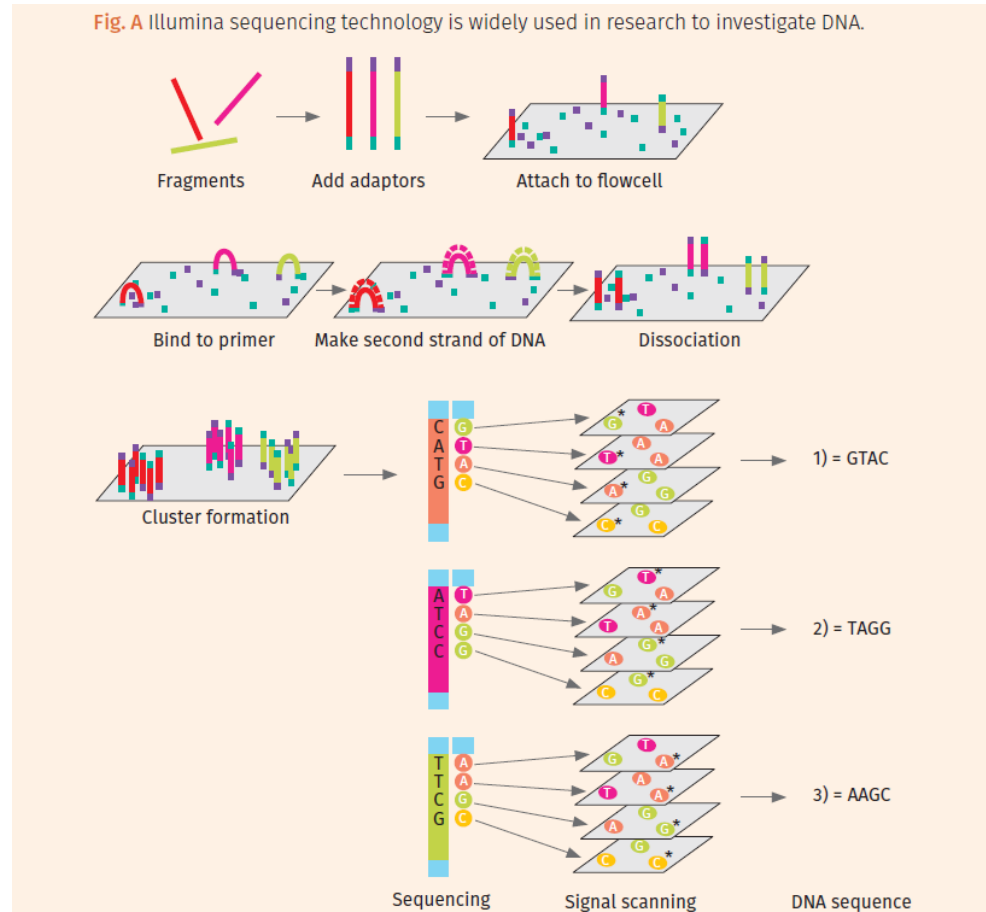
HiSeq Flow Cells



Illumina uses a glass ‘flowcell’, about the size of a microscope slide, with 8 separate ‘lanes’.

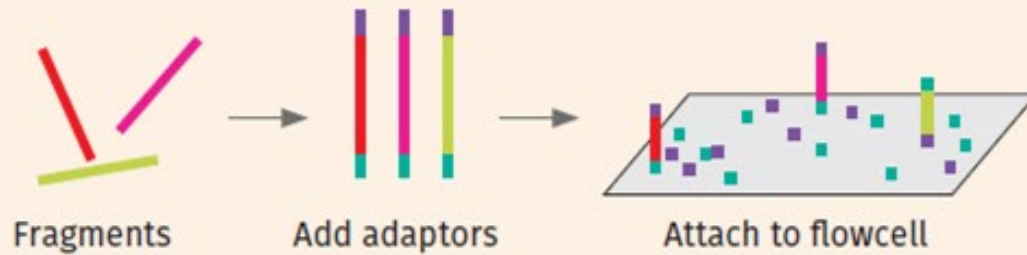
The HiSeq instrument scans both upper and lower surfaces of each flowcell lane.

Illumina sequencing: overview

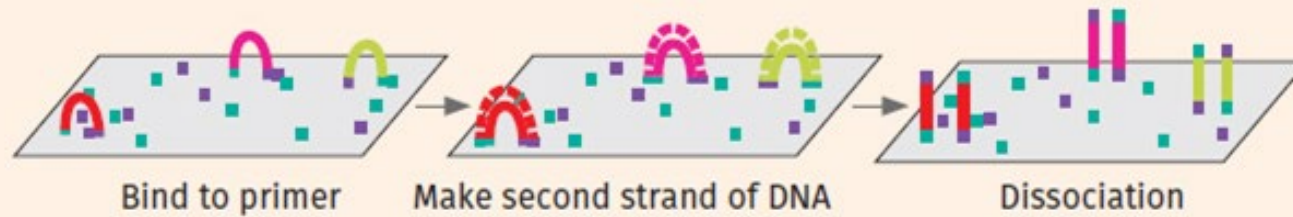


“Seeding” the flow cell

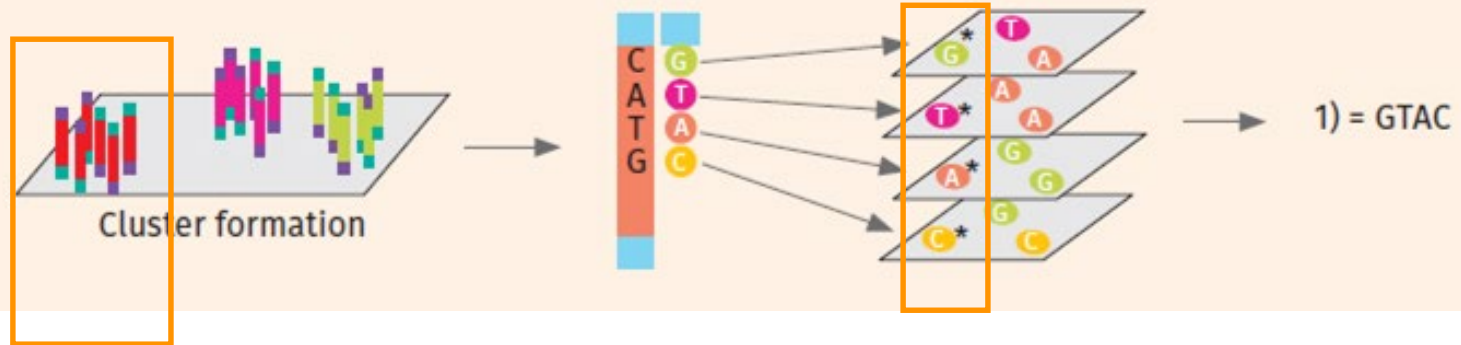
Fig. A Illumina sequencing technology is widely used in research to investigate DNA.



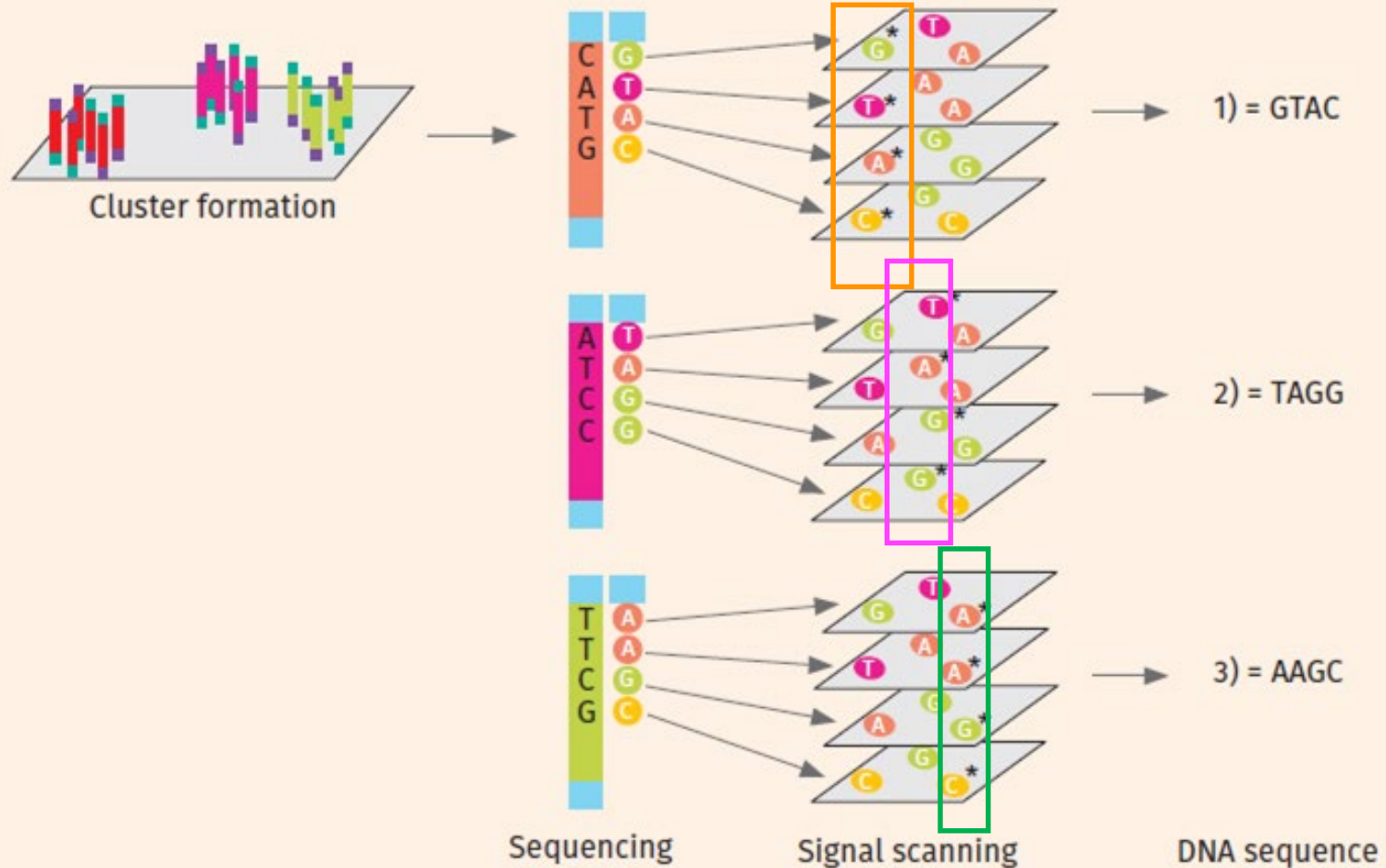
Forming clusters



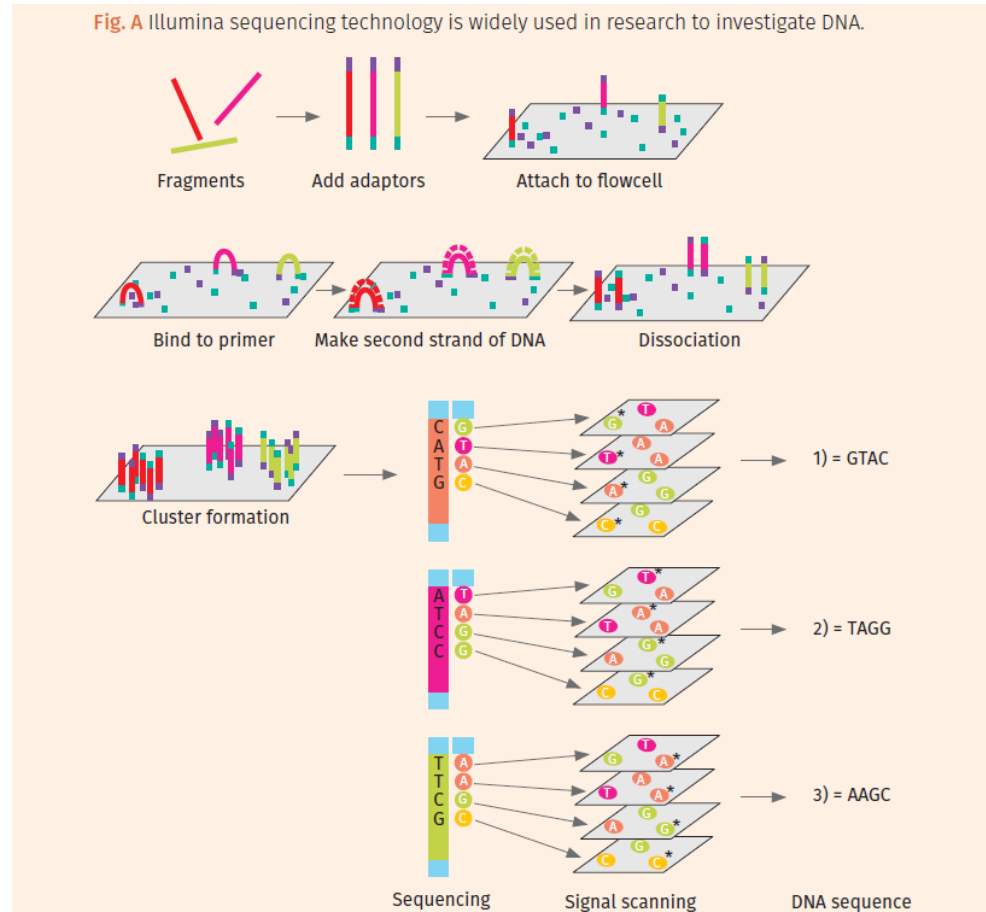
Reading each cluster



Reading each cluster



Illumina sequencing: overview



Data analysis: lots of data!



Sequencing in remote environments: possible?



Oxford Nanopore



Oxford Nanopore

